**Functional Characterization of the Infection-Inducible Peptide Edin in Drosophila melanogaster**

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**Abstract**

Drosophila is a well-established model organism for studying innate immunity because of its high resistance against microbial infections and lack of adaptive immunity. In addition, the immune signaling cascades found in Drosophila are evolutionarily conserved. Upon infection, activation of the immune signaling pathways, Toll and Imd, leads to the expression of multiple immune response genes, such as the antimicrobial peptides (AMPs). Previously, we identified an uncharacterized gene edin among the genes, which were strongly induced upon stimulation with Escherichia coli in Drosophila S2 cells. Edin has been associated with resistance against Listeria monocytogenes, but its role in Drosophila immunity remains elusive. In this study, we examined the role of Edin in the immune response of Drosophila both in vitro and in vivo. We report that edin expression is dependent on the Imd-pathway NF-κB transcription factor Relish and that it is expressed upon infection both in vitro and in vivo. Edin encodes a pro-protein, which is further processed in S2 cells. In our experiments, Edin did not bind microsor, nor did it possess antimicrobial activity to test microbial strains in vitro or in vivo. Furthermore, edin RNAi did not significantly affect the expression of AMPs in vitro or in vivo. However, edin RNAi flies showed modestly impaired resistance to E. faecalis infection. We conclude that Edin has no potent antimicrobial properties but it appears to be important for E. faecalis infection via an uncharacterized mechanism. Further studies are still required to elucidate the exact role of Edin in the Drosophila immune response.

**Introduction**

Innate immunity is the first line of defense in all multicellular organisms. During the last few decades, the fruit fly *Drosophila melanogaster* has proven to be well suited for studying innate immune responses. In contrast to vertebrates, *Drosophila* only has an innate immune system, which is highly sophisticated and in part conserved among higher organisms [1]. In *Drosophila*, effective innate immune responses are based on the ability of several pattern-recognition receptors to recognize and bind common microbial surface structures. One main outcome of this initial microbial recognition is the activation of NF-κB immune signaling pathways, which leads to the production of several potent antimicrobial peptides (AMPs).

In *Drosophila*, the production of AMPs is mainly regulated by two NF-κB signaling pathways: the Imd (immune deficiency) pathway [2] reviewed in [3] and the Toll pathway [4] reviewed in [5]. Both of these pathways are highly conserved from fly to man. The Imd pathway is activated by dianaminopimelic acid-type peptidoglycan (DAP) [6], present in most or all Gram-negative bacteria, but also in some Gram-positive bacteria like *Listeria monocytogenes*. The Toll pathway is activated mainly by the lysine-type peptidoglycan present in many other Gram-positive bacteria [7], reviewed in [5]. Both of these signaling pathways can also be induced by different fungi [8,9]. Activation of the Imd and Toll signaling pathways upon microbial infection ultimately causes the nuclear translocation of the NF-κB transcription factors, Relish or Dif/Dorsal respectively, leading to the expression of dozens of NF-κB responsive genes [10,11,12,13,14]. The molecular function of many of these genes still remains unknown.

Earlier, we identified a gene, *CG32185*, to be highly induced in S2 cells in response to heat-killed *Escherichia coli* [14]. Later, Gordon et al. called the gene *edin* and found it to be associated with *Listeria monocytogenes* resistance [15]. In addition, it has been shown that Edin is secreted into the hemolymph in *Drosophila* third instar larvae upon infection [16]. Because the molecular function of Edin and the signaling pathways involved are still mainly unknown, in our current study we set out to examine the role of Edin in the *Drosophila* immune response both in vitro and in vivo.
Results

Edin expression is Relish-dependent in vitro and in vivo upon Gram-negative bacterial infection

When Drosophila encounters microbes, several signaling pathways are activated leading to transcriptional modifications. This response varies depending on the microbe and the site of infection. During a systemic infection, the expression of dozens of genes is induced [11,12] leading to very effective defense responses. Upon infection, most of the highly induced genes are known to be AMP genes, DIsMs (Drosophila immune-induced molecules) or genes related to signal regulation. Nevertheless, the molecular function of several of the induced genes is yet to be characterized. Previously, we studied which genes are induced in response to heat-killed Escherichia coli in Drosophila macrophage-like S2 cells [14]. Table I represents the oligonucleotide microarray data of the most strongly induced genes (data collected from [14]). The eight most strongly induced genes encode five known AMPs, one peptidoglycan recognition protein (PGRP-LB), a negative regulator of the IMD pathway (pik) [17] and Edin (CG32185). According to the microarray results, the expression of Edin is strongly induced within hours after the bacterial challenge and the induction pattern of Edin resembles that of known antimicrobial peptides (Table 1).

In S2 cells, the response to E. coli is known to be predominantly mediated via the IMD pathway [15]. To verify whether the induction of Edin is dependent on the IMD pathway, we silenced the IMD pathway by knocking down the transcription factor Relish by RNAi. The induction of Edin was completely abolished in Relish dsRNA treated S2 cells at the 4 h time point (Table I) indicating that Edin expression is regulated via the IMD pathway in S2 cells after induction with heat-killed E. coli.

The edin gene encodes a short peptide of 115 amino acids including an N-terminal signal sequence (amino acids 1–22) (Figure 1A). The predicted signal peptide cleavage site is supported by proteomic data from Verleyen et al. [16], who identified the predicted amino terminal of the mature protein in peptide fragments from hemolymph. Likely orthologs of the edin gene can be found in other brachycerean flies, including all sequenced Drosophila species, but not in other insects (Figure 1A). For Musca domestica, three isoforms are represented in the EST databases (not shown). A tendency for pseudogenisation of the edin gene can be noted, as stop codons are present in the D. yakuba and D. mojavensis homologs. For the latter, an apparently functional gene can be noted, as stop codons are present in the Drosophila edin homologs. For the latter, an apparently functional gene can be noted, as stop codons are present in the Drosophila edin homologs.

Table 1. Induction of Drosophila antimicrobial peptide genes and edin in E. coli -challenged S2 cells (data collected from [14]).

<table>
<thead>
<tr>
<th>Gene</th>
<th>#CG</th>
<th>0 h</th>
<th>0.5 h</th>
<th>1 h</th>
<th>4 h</th>
<th>24 h</th>
<th>Relish RNAi 4 h</th>
</tr>
</thead>
<tbody>
<tr>
<td>Attacin B</td>
<td>CG18372</td>
<td>1±0.1</td>
<td>1.5</td>
<td>6.0</td>
<td>60.6±15.1</td>
<td>87.2±87.2</td>
<td>0.1±0.0</td>
</tr>
<tr>
<td>Diptericin B</td>
<td>CG10794</td>
<td>1±0.0</td>
<td>2.3</td>
<td>3.6</td>
<td>52.4±4.3</td>
<td>78.1±1.8</td>
<td>0.2±0.1</td>
</tr>
<tr>
<td>Attacin D</td>
<td>CG7629</td>
<td>1±0.0</td>
<td>1.1</td>
<td>2.6</td>
<td>47.5±6.3</td>
<td>92.5±2.1</td>
<td>0.1±0.1</td>
</tr>
<tr>
<td>Metchnikowin</td>
<td>CG8175</td>
<td>1±0.0</td>
<td>1.7</td>
<td>6.5</td>
<td>41.3±15.7</td>
<td>52.2±2.0</td>
<td>0.4±0.1</td>
</tr>
<tr>
<td>Edin</td>
<td>CG32185</td>
<td>1±0.1</td>
<td>0.9</td>
<td>3.5</td>
<td>29.8±6.4</td>
<td>48.5±1.1</td>
<td>0.0±0.0</td>
</tr>
<tr>
<td>Pik</td>
<td>CG15678</td>
<td>1±0.2</td>
<td>2.0</td>
<td>15.5</td>
<td>15.1±0.6</td>
<td>5.4±0.0</td>
<td>0.4±0.1</td>
</tr>
<tr>
<td>PGRP-LB</td>
<td>CG14704</td>
<td>1±0.0</td>
<td>1.2</td>
<td>2.0</td>
<td>8.5±2.0</td>
<td>20.7±0.3</td>
<td>0.7±0.2</td>
</tr>
<tr>
<td>Cecropin B</td>
<td>CG1878</td>
<td>1±0.0</td>
<td>1.2</td>
<td>3.2</td>
<td>7.4±0.9</td>
<td>4.0±0.3</td>
<td>0.6±0.0</td>
</tr>
</tbody>
</table>

Iterated PSI-BLAST searches indicate that Edin is related to the Attacin/Diptericin superfamily of glycine-rich antibacterial peptides. The best hits were to Drosophila circulus Dipterocin B (E = 8e-20) and Hyalophora cecropia Attacin E (E = 2e-18). Figure 1A shows an alignment to Dipterocin B and the C-terminal (G2) domain of Attacin A from D. melanogaster.

Since Edin has a predicted signal sequence, we next examined if Edin is actually secreted from cells. To test this, we cloned edin cDNA into the heavy metal-inducible expression vector pMT/V5, transfected S2 cells with the construct and analyzed the presence of the protein both in the cell culture medium and cell extracts by western blotting using an anti-V5 antibody. In the S2 cells, both shorter and longer forms of Edin were detected, corresponding to V5-tagged peptides with and without the signal sequence, respectively. In the cell culture medium, only the shorter, C-terminal form, without the signal sequence could be observed (Figure 1B). This result suggests that Edin has a functional signal sequence, which is cleaved before the peptide is secreted. These results are in line with the report of Verleyen and coworkers [16], who detected amino-terminal fragments of Edin with mass spectrometry in the hemolymph of Drosophila larvae infected with a mixture of Gram-negative and Gram-positive bacteria.

Since the expression of Edin is Relish-dependent in vitro, we next investigated whether edin is also induced upon microbial challenge in vivo. We infected wild-type Canton S and Relish null mutant adult flies (RelE20) with the Gram-negative bacteria, Escherichia coli. Total RNAs were extracted and the transcript levels of edin were determined with RT-PCR and agarose gel electrophoresis. As shown in Figure 1C, edin is induced in Canton S but not in RelE20 mutant flies. Attacin A was used as a positive control and showed a similar expression pattern to edin (Figure 1C). These results together with the previously published microarray data indicate that edin expression is strongly and rapidly induced upon a Gram-negative bacterial infection in a Relish-dependent manner both in vitro and in vivo. These results together propose that Edin has a function related to microbial resistance. Thus, we next subjected Edin to further functional characterization both in vitro and in vivo.

Edin has no significant effect on bacterial binding

The phagocytosis of invading microbes is an essential component of Drosophila immunity [18,19]. To this end we tested whether Edin has a role in bacterial binding or opsonization. Plasmatocyte-like S2 cells that are capable of binding and phagocytosing microbes [20] were treated with edin dsRNA and the ability of the cells to bind heat-killed, fluorescently labeled E. coli and Staphylococcus aureus was analyzed by flow cytometry. As a positive control, we used a dsRNA treatment targeting eater, which
codes for an important phagocytic receptor for bacteria both in S2 cells and in *Drosophila in vivo* [18,19,21]. GFP dsRNA was used as a negative control. *Edin* RNAi did not affect the ability of S2 cells to bind *E. coli* (Figure 2A). Likewise, *edin* dsRNA treatments did not compromise the ability of S2 cells to bind *S. aureus* (Figure 2B) but rather seemed to modestly enhance the binding activity of S2 cells.

To test the effect of *edin* overexpression on bacterial binding, S2 cells were first transiently transfected with a pMT-*edin*V5 construct. An empty pMT/V5 plasmid was transfected as a control. 24 h after transfection, CuSO₄ was added to the cell culture medium to induce the expression of the construct. Two days later, the medium was collected and transferred to other S2 cells which were pre-treated with *edin* dsRNA to block endogenous *edin* expression. Thereafter, FITC-labeled, heat-killed *E. coli* or *S. aureus* were added and the amount of cell-associated bacteria was monitored using flow cytometry. In line with the results of *edin* overexpression, *edin* RNAi did not affect the ability of S2 cells to bind *E. coli* (Figure 2A). Likewise, *edin* dsRNA treatments did not compromise the ability of S2 cells to bind *S. aureus* (Figure 2B) but rather seemed to modestly enhance the binding activity of S2 cells.

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RNAi experiments, edin overexpression had no effect on the binding of E. coli (Fig. 2C) or S. aureus (Fig. 2D). The presence of Edin in the cell-culture medium was confirmed by western blotting using an anti-V5 antibody (data not shown).

To investigate in a more direct way if Edin binds microbes, we incubated Edin-containing cell culture medium with live E. coli, Serratia marcescens, Staphylococcus epidermidis, Enterococcus faecalis, Listeria monocytogenes, Micrococcus luteus, Saccharomyces cerevisiae and S. aureus. Latex beads (carboxylated polystyrene), which are expected to bind all kinds of proteins to some extent, were used as a positive control. The microbial suspensions were incubated with 500 μl of Edin-containing medium at +4°C after which the microbes were pelleted and washed with PBS. Finally, the pellets were suspended and boiled in an SDS-PAGE sample buffer to detach bound Edin from the microbes before electrophoresis. Next, the proteins were transferred onto nitrocellulose membranes and Edin was detected using an anti-V5 antibody. As a reference, 20 μl of Edin-containing medium was loaded into the first lane. Therefore, if Edin attached efficiently to the indicated microbe, much more Edin should be detected in the samples (500 μl Edin-containing medium used) compared to the reference lane (20 μl Edin-containing medium). As shown in Figure 3 (the rightmost lanes), carboxylated latex beads, i.e. the positive control, bound Edin. In contrast, virtually no Edin was bound to the tested Gram-negative bacteria, E. coli and S. marcescens. Furthermore, only a faint signal was detected with the Gram-positive bacteria S. epidermidis, E. faecalis, L. monocytogenes, M. luteus and S. aureus, and with the baker’s yeast S. cerevisiae as compared to the reference lane (ctrl in Figure 3). Based on these results, we conclude that Edin does not strongly bind any of the tested microbes.

The effect of Edin on immune signaling

Next, we investigated whether Edin is involved in modulating the activity of Drosophila innate immune signaling cascades. S2 cells were transfected with luciferase-reporter constructs together with edin dsRNA as well as with negative and positive control dsRNAs, and the luciferase activities of the cell lysates were analyzed. Transfection efficacy and cell viability were assessed with an Actin-5C-galactosidase reporter. GFP dsRNA was used as a negative control in all assays. First, we tested the effectiveness of edin RNAi in vitro by treating S2 cells with GFP or edin dsRNAs, and analyzing the relative expression levels of edin. As shown in Figure 4A, edin RNAi abolishes the endogenous edin expression.

In order to analyze the Imd pathway activity, an Attacin A-luciferase reporter and Relish dsRNA as a positive control were used and the pathway was activated by adding heat-killed E. coli to the cell culture medium. The samples were collected 0 h (no induction), 1 h, 4 h, 8 h and 24 h after E. coli induction. As expected, Relish RNAi strongly decreases the Imd-pathway activity at all time points (Figure 4B). On the contrary, edin RNAi had minor or no effect in this setting, although at the 24 h time point there was a trend for reduced Attacin A promoter driven luciferase activity.
Edin has no potent antimicrobial properties in vitro or in vivo

The kinetics of edin expression closely resembles those of known AMP genes, which led us to examine whether Edin has antimicrobial properties in vitro or in vivo. To study this, we first analyzed whether Edin was able to limit bacterial growth in vitro. We overexpressed edin in S2 cells, collected the cell culture medium and incubated the medium either with E. coli or S. aureus. Medium from S2 cells transfected with an empty vector was used as a control. As shown in Figure 6A and 6B, E. coli and S. aureus grew equally well in control medium and in medium containing Edin.

To further investigate the antimicrobial properties of Edin in vitro, we designed synthetic peptides containing the amino acids 22–45 (Edin C-terminal form) or 50–115 (Edin N-terminal form). The peptides were tested for their ability to reduce bacterial growth in vitro. Cecropin A and Lysozyme were used as positive controls for Gram-negative and Gram-positive bacteria, respectively. The peptides were incubated with E. coli (Figure 6C–D), E. cloacae (Figure 6E–F), L. monocytogenes (Figure 6G–H) or E. faecalis (Figure 6I–J) and colony forming units were determined. As shown in Figure 6C–J, Cecropin A and Lysozyme at their highest concentrations almost abolished the growth of the tested microbes whereas neither the synthetic C-terminal or N-terminal form of Edin was able to affect the growth of the bacteria. Moreover, no synergistic effects were observed when Edin was incubated together with either Cecropin A or Lysozyme (three rightmost columns in Figure 6 panels C–J).

To test the antimicrobial properties of Edin in a more physiological context, the effect of edin overexpression on the survival of flies after bacterial infections was analyzed. First, to test whether overexpressing edin affects survival or lifespan, the UAS-edin,RelE20 overexpression line was crossed with the Act5C-GAL4/CyO driver line and the lifespan of the offspring was monitored. As shown in Figure 7A, overexpression of edin did not affect the old offspring by septic injury with E. cloacae. Flies crossed with w^1118 flies were used as controls. As shown in Figure 5A, in vivo RNAi of edin using the C564-GAL4 driver strongly suppresses edin expression in whole flies, indicating that the UAS-RNAi construct is effective. UAS-edin,RelE20 flies crossed with the C564-GAL4 driver showed expression levels comparable to the E. cloacae infected control flies (Figure 5B).

In agreement with our in vitro results, in vivo RNAi of edin did not show any clear effect in the expression levels of the tested AMP genes (two left-most panels, Figure 5C–H). There is a trend towards a minor decrease at the 4 h time points of the tested AMPs, excluding Drosocin (Figure 5H), but the decrease was statistically significant only with Cecropin A1 (Figure 5D) and Attacin B (Figure 5E). We next tested whether overexpression of edin affects the production of AMPs via the Imd pathway. We compared AMP expression after septic injury with E. cloacae between UAS-edin flies crossed with C564-GAL4 and UAS-edin flies crossed with w^1118 flies. We observed moderate increase only in Drosocin expression at the 8 h time point (68% increase for p<0.05) (Figure 5H). Noteworthy, edin expression did not activate AMP gene expression without a microbial challenge (see the 0 h time point in the rightmost panel in Figure 5C–H). This is in line with the results in S2 cells and rules out the possibility that Edin would function as a cytokine mediating immune response from the site of induction to other tissues (for example from hemocytes to the fat body). Based on these results, we conclude that Edin has no important role in the regulation of the Imd pathway activity either in vitro or in vivo.

Edin activity (Figure 4B). Because edin RNAi appeared to have a minor effect on the Imd pathway activity when induced with heat-killed E. coli at the 24 h time point, we next investigated the effect of the edin dsRNA with other pathway elicitors. To this end, heat-killed S. marcescens, heat-killed E. cloacae, peptidoglycan and overexpression of the cytoplasmic tail of the PGRP-LC receptor were used. As shown in Figure 4C, edin RNAi had no effect on the Akt-luciferase activity in this experimental setting. These results indicate that Edin does not have an important role in the regulation of the Imd pathway activity in S2 cells.

To investigate the role of Edin in the Toll pathway signaling, we used a Drosomycin-luciferase reporter, and MyD88 dsRNA as a positive control, and activated the pathway by transfecting the cells with a constitutively active form of the Toll receptor, Toll^Act5C (Figure 4D) or with the cleaved, active Spätzle ligand (Figure 4E). For the JAK/STAT signaling pathway, we used TurandotM-luciferase reporter and STAT dsRNA as a positive control (Figure 4F). The pathway was activated by overexpressing hopscotch (Tum)^2, the active form of Drosophila Jak. Edin RNAi did not significantly affect the signaling via the Toll pathway (Figure 4D–E), or the JAK/STAT pathway (Figure 4F). These results indicate that Edin has no central role in regulating immune signaling in vitro.

To test the role of Edin in Imd pathway regulation in vivo, we monitored the Imd pathway-mediated AMP gene expression levels with qRT-PCR in edin RNAi flies and in edin overexpression flies we created. The overexpression flies were created by microinjecting the pUAST-edin construct into RelE20 mutant embryos. To analyze Imd pathway activity, edin RNAi (VDRG #14209) and UAS-edin,RelE20 flies were crossed with the C564-GAL4 driver that targets transgene expression to the fat body in addition to some other organs [22]. The Imd pathway was then activated in week-old offspring by septic injury with E. cloacae. Flies crossed with w^1118 flies were used as controls. As shown in Figure 5A, in vivo RNAi of edin using the C564-GAL4 driver strongly suppresses edin expression in whole flies, indicating that the UAS-RNAi construct is effective. UAS-edin,RelE20 flies crossed with the C564-GAL4 driver showed expression levels comparable to the E. cloacae infected control flies (Figure 5B).

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lifespan of the flies and was comparable to that of the control flies. Furthermore, \textit{edin} expression did not compromise the development of flies since equal amounts of \textit{UAS-edin,RelE20/ActGAL4} and \textit{UAS-edin,RelE20/CyO} flies were obtained from the crosses (Figure 7B). Similar results were obtained when \textit{edin} overexpression flies were crossed with either the \textit{C564-GALA} driver line or the ubiquitous \textit{daughtersless-GALA} driver line (data not shown).

An earlier study has shown that the expression of a single AMP can restore antimicrobial activity in \textit{Drosophila} [23]. To test whether the expression of \textit{edin} is sufficient to enhance resistance against septic infection in adult flies, we expressed \textit{edin} in a...
Figure 5. The effect of Edin on AMP production in vivo. Edin RNAi and overexpression flies (edin,Re^{F20}) were crossed with C564-GAL4 flies or w^{1118} flies as a control, their offspring was infected with E. cloacae, total RNAs were extracted at indicated time points and qRT-PCR for the indicated genes was performed. (A) Expression of edin is knocked down in edin RNAi flies crossed to C564-GAL4 driver flies. (B) Edin overexpression flies express edin at a physiological level. Edin overexpression flies crossed with C564-GAL4 have slightly higher levels of edin compared to flies crossed with w^{1118}. For (A–B) the data are pooled from 2 independent experiments, and n = 8 for each sample at each time point. (C–H) The effect of edin RNAi and
homzygous RelR^200 mutant background using a C564-GAL4;RelR^200 line. In the homzygous RelR^200 background, AMP production via the IMD pathway is eliminated making the flies very sensitive to infections with Gram-negative bacteria [24]. To test whether Edin had antimicrobial properties against Gram-negative or Gram-positive bacteria in vivo, we infected the UAS-edin, RelR^200 flies crossed with the C564-GAL4,RelR^200 driver with the Gram-positive bacterium L. monocytogenes (Figure 7C), which has a DAP-type peptidoglycan, with the Gram-negative bacterium E. faecalis (Figure 7D), and with the Gram-positive bacterium E. cloacae (Figure 7E). In this homzygous RelR^200 background, overexpression of edin did not affect the survival rate upon septic injury with any of these microbes. In addition, no rescue was observed after a septic E. coli infection (data not shown). According to the results, edin overexpression was not sufficient to rescue the flies from succumbing to bacterial infection (Figure 7C-E) indicating that Edin alone does not possess sufficient antimicrobial properties against Gram-negative or Gram-positive bacteria.

To test whether Edin has antimicrobial properties in the context of a normal functioning immune response in Drosophila, we overexpressed edin in a heterozygous RelR^200 mutant background. Edin overexpression flies crossed with C564-GAL4 were infected with L. monocytogenes (Figure 7F), E. cloacae (Figure 7G) and E. faecalis (Figure 7H) and monitored for survival. As shown in Figure 7F–H, overexpressing edin did not protect the flies from the bacterial infection. Together these results indicate that Edin has no antimicrobial properties against either Gram-negative or Gram positive bacteria in vitro or in vivo. These results argue that Edin has another immune response modulating function.

Edin is required for normal resistance against bacteria

Next, we investigated whether Edin is required for normal resistance against septic infection. To this end edin RNAi flies were crossed with the C564-GAL4 driver or w^1118 flies as a control, and the one-week-old offspring were infected with E. cloacae, E. faecalis or L. monocytogenes, RelR^200 mutant flies were used as a positive control in the E. cloacae and L. monocytogenes infection model, and UAS-MyD88 RNAi flies crossed with the C564-GAL4 driver as a positive control in the E. faecalis infection model. When infected with the Gram-negative bacterium E. cloacae, RelR^200 mutant flies succumbed to the infection within 24 h. Edin RNAi flies crossed with C564-GAL4 flies showed a mild decrease in survival after E. cloacae infection compared to edin RNAi flies crossed with w^1118 (Figure 9A) but this it not significant because the C564-GAL4 driver flies crossed to w^1118 are more susceptible to the infection. However, a decrease in survival was observed in edin RNAi flies infected with the Gram-positive bacterium E. faecalis (Figure 8B). However, no statistically significant difference in survival was seen after an L. monocytogenes infection (Figure 8C), although a similar trend in survival could be observed, which is in line with the results by Gordon et al. [15]. These results imply that the expression of edin might be required for normal resistance against some bacterial infections.

Discussion

In Drosophila, the expression of many genes is induced in response to microbial infection. In this study, we examined the role of the infection-inducible gene edin in the immune response of Drosophila melanogaster both in vitro and in vivo. We show that edin is highly induced in S2 cells by E. coli and its expression is dependent on the NF-kB transcription factor Relish both in vitro and in vivo. In line with the results of Verleyen and coworkers [16], we observe that Edin has a functional signal sequence leading to its cleavage and secretion from S2 cells. Despite the fact that edin is highly induced upon infection and that its expression pattern resembles that of known AMPs, we were not able to observe any antimicrobial properties in vitro or in vivo. Nor were we able to see any bacterial binding or opsonization when these properties of Edin were studied. Edin expression also was dispensable for AMP expression via the IMD pathway both in vitro and in vivo. However, interestingly edin RNAi flies showed decreased survival after bacterial infection with E. faecalis.

Traditionally, most studies on Drosophila AMPs have been successfully carried out in vitro. However, Drosophila is also a powerful model system for studying the activity of antimicrobial peptides in vivo, since it is easy to produce immunocompromised mutant fly lines, which are viable and fertile. Earlier studies have shown that Drosophila mutants of the Toll and IMD pathway, that have impaired production of AMPs via these signaling pathways, are highly susceptible to microbial infections [2,4,24] and even a single bacterial cell can be enough to kill a mutant fly [24]. The antimicrobial properties and the microbial specificity of a gene product can be studied by overexpressing the gene of interest in the mutant background of choice. It has been reported that the overexpression of a single antimicrobial peptide in Toll and IMD pathway double mutant flies can restore the resistance to a microbial infection to a level comparable to that of wild-type flies [23]. In our current study, we were not able to demonstrate a broad antimicrobial role for Edin in vitro or in vivo. In vitro, we observed no effect on the colony forming of bacterial cells when Edin was produced in S2 cells or when synthetic peptides were used.

In vivo, the effect of edin overexpression on the resistance against microbial infection was analyzed both in a homozygous RelR^200 mutant background and in a heterozygous background. RelR^200 mutants were selected since they are highly sensitive to Gram-negative bacterial infections. However, no increase in survival after septic injury could be observed in either one of these backgrounds. Therefore it is likely that Edin does not have an antimicrobial role in Drosophila although it is highly expressed upon bacterial infection. However, it is also possible that Edin is effective only against a specific microbe which we did not test in our current study. The in vivo analysis of antimicrobial properties of a certain peptide is further complicated by the production of a large battery of AMPs that can be partially redundant in their specificities. For instance, Edin alone might not be sufficient to fight against microbial infections, but it may require the presence of another AMP or other immune effector molecules, for full activity.

Previously, Gordon and coworkers [15] have reported that high expression levels of edin are detrimental to fly survival and lifespan. We carried out lifespan experiments with our edin overexpression fly line and analyzed the proportions of the closed progeny. In contrast to Gordon et al., we did not observe a negative effect of edin overexpression on fly survival or lifespan. This difference in results could be due to different expression levels of edin or different genetic backgrounds of the flies used in these studies. According to our results, the edin overexpression fly line used in this study shows expression levels comparable to expression levels upon septic infection (Figure 5B). Furthermore, Gordon et al. [15] reported
Figure 6. Edin has no broad antimicrobial properties against Gram positive or Gram negative bacteria in vitro. (A–B) Edin does not limit the growth of *E. coli* or *S. aureus* in S2 cell culture medium. S2 cells were transfected with a copper-inducible pMT-edin-V5 or an empty pMT vector, and the abilities of *E. coli* and *S. aureus* to proliferate in these mediums were analyzed. (C–G) Synthetic forms of Edin do not limit the growth of *E. coli* (C), *E. cloacae* (D), *L. monocytogenes* (E), *E. faecalis* (F) or *S. aureus* (G). Both N-terminal and C-terminal forms of Edin were tested. Bacteria were cultured to an OD600 nm of 0.33, incubated with synthetic Edin and the ability of the bacteria to grow was analyzed. Cecropin A and Lysozyme were used as positive controls for Gram-negative and Gram-positive bacteria, respectively. Left column, N-terminal Edin; right column, C-terminal Edin. doi:10.1371/journal.pone.0037153.g006
that Edin is required for resistance against *Listeria monocytogenes* infections. *L. monocytogenes* is a DAP-type peptidoglycan containing intracellular bacterium which can infect both mammals and *Drosophila* [25,26]. Gordon et al. [15] report a significant decrease in survival after *L. monocytogenes* infection with two independent *edin* RNAi lines indicating that the normal *edin* expression is required...
We also analyzed the role of Edin as a modulator of innate immune signaling cascades. Nevertheless, our experiments indicate that Edin no strong effect on Imd pathway activity either in vitro or in vivo.

We conclude that the expression of edin is Relish-dependent both in vitro and in vivo but further studies are required to elucidate the exact role of Edin in the immune response in *Drosophila*. Also the mechanisms and signaling pathways involved in the *Listeria monocytogenes* infection remain to be studied.

### Materials and Methods

**Oligonucleotide microarrays**

Oligonucleotide microarray expression data of S2 cells was collected from [14].

**Microbial culture**

*Listeria monocytogenes* (strain 10403S), *Enterococcus faecalis*, *Staphylococcus aureus* and *Staphylococcus epidermidis* were cultured in BHI. *Enterobacter cloacae* (strain B12) and *Micrococcus luteus* were cultured in LB supplemented with either 15 ng/ml of nalidixic acid (Sigma-Aldrich, St. Louis, Missouri, USA) or 100 µg/ml of streptomycin (Sigma-Aldrich), respectively. *Serratia marcescens* (strain Db11) and *Escherichia coli* were cultured in LB supplemented with 100 µg/ml of ampicillin. The baker’s yeast *Saccharomyces cerevisiae* (AH109) was grown overnight inYPD medium (Gibco/Life Technologies, Carlsbad, CA, USA) supplemented with 15 µg/ml of kanamycin at +30°C with shaking.

**Semi-quantitative and quantitative RT-PCR**

Semi-quantitative RT-PCR reactions for *edin*, *Attacin A* and *Act5C* were performed using Super-Script™ II One-Step RT-PCR with Platinum Taq kit (Invitrogen/Life Technologies, Carlsbad, CA, USA). The following primers were used: *Edin*: 5′-GTTCTCCAA-CAAGTGCGG-3′ (forward), and 5′-CAGAAATGCGAGG-TGCCC-3′ (reverse); *Attacin A*: 5′-TTTGCGCTACAACAAATTGCTG-3′ (forward), and 5′-GGCTTCGTTGTTGCAACAGG-3′ (reverse); *Act5C*: 5′-CGAAGATGTTGCTGCTTG-3′ (forward), and 5′-AGAACATCCGTTGTA-3′ (reverse).

Quantitative RT-PCR was carried out using the QuantiTect SYBR Green RT-PCR kit (Qiagen) and an ABI7000 (Applied Biosystems) instrument according to the manufacturer’s instructions. Results were analyzed with the ABI 7000 System SDS software version 1.2.3. The following primers were used: *Atth*: 5′-CAGTTTCACACAGGAC-3′ (forward) and 5′-CTCC-CTGGTGAAAGC-3′ (reverse); *Drosocin*: 5′-TTTCTCGTGTGGTTC-3′ (forward) and 5′-TGCCGCGT-3′ (reverse); *Atkt*: 5′-GATCGTACCATGTCCGC-3′ (forward); *AttA*: 5′-CATCGTATGGGTGATG-3′ (forward) and 5′-GGATCG-3′ (reverse); *Act5C*: 5′-CATCGTATGGGTGATG-3′ (forward) and 5′-GGATCG-3′ (reverse); *Act5C*: 5′-CATCGTATGGGTGATG-3′ (forward) and 5′-GGATCG-3′ (reverse); *GcCl*: 5′-CATCGTATGGGTGATG-3′ (forward) and 5′-GGATCG-3′ (reverse); *Dpdb*: 5′-GATCGTATGGGTGATG-3′ (forward) and 5′-GGATCG-3′ (reverse); *Act5C*: 5′-CATCGTATGGGTGATG-3′ (forward) and 5′-GGATCG-3′ (reverse); *Act5C*: 5′-CATCGTATGGGTGATG-3′ (forward) and 5′-GGATCG-3′ (reverse); *Act5C*: 5′-CATCGTATGGGTGATG-3′ (forward) and 5′-GGATCG-3′ (reverse); *Act5C*: 5′-CATCGTATGGGTGATG-3′ (forward) and 5′-GGATCG-3′ (reverse).

**S2 cell culture and transfections**

*Drosophila* hemocyte-like S2 cells [27] (obtained from Invitrogen/Life Technologies) were maintained in Schneider’s Insect Cell Culture Medium (Sigma-Aldrich, St. Louis, Missouri, USA) supplemented with 10% FBS, 100 U/ml Penicillin and 100 µg/ml Streptomycin (Sigma-Aldrich) at +25°C. The cells were transfected using the Fugene® transfection reagent (Roche Applied...
Science, Penzberg, Germany) according to the manufacturer’s instructions.

**Cloning and constructs**

*Edin* was cloned into the pMT/V5-HisA (Invitrogen/Life Technologies) and pUAST [28] vectors using S2 cell cDNA as a template. The primers used were 5'-CAGAATTCTAGTTCTC-CACACAGTGCC-3' and 5'-CAGGGTACCTCAGAAATGCCA-GGTGCCC-3' for pUAST, and 5'-CAGGCGGCGCATG-TTCTCCAAACAGTGCC-3' and 5'-CAGCCAGGAATGCCA-GGTGCCCAGG-3' for pMT/V5-His.

**Western blotting**

S2 cells were transfected with 0.5 μg of pMT-[*edin*]-V5. Cells were harvested, pelleted and lyzed 24 h after addition of CuSO4. 25 μg of cell lysate and supernatant were electrophoresed in NuPAGE 12% Bis-Tris gel (Invitrogen Life Technologies), blotted on a nitrocellulose membrane, and detected by Western blotting using mouse anti-V5 primary Ab (Invitrogen/Life Technologies) and goat anti-mouse Ab HRP conjugates (Molecular Probes) together with ECL Plus Western blotting detection system (GE Healthcare Life Sciences, Uppsala, Sweden).

**Synthetic peptides**

Two forms of synthetic Edin were ordered from Peptide 2.0 (Chantilly, VA, USA). Amino acid sequences: N-terminal form, SYRQ PYPEEF QTSPQ LQGLV YAVPLV; C-terminal form, SPEGG SVVVT ASKDNQ VGREAS VQYNHN LYSYG DGRGS IDAYA QASRN FDYNR NNVEG GIRGT WHF. The peptides were dissipated in H2O according to the manufacturer’s instructions.

**Colonies forming unit assay**

Edin-V5 expressed in S2 cells: S2 cells in 48-well plates in an antibiotic-free medium were transfected with 0.5 μg of pMT-[*edin*-V5 plasmid or an empty plasmid. Expression of the plasmid was used as a positive control for Gram-positive and Gram-negative strains. Serial dilutions were made in sterile water. Dilutions were 25, 100 nM. Suspensions were incubated for 2 h at 37°C.

**Luciferase reporter assays and dsRNA treatments**

Luciferase reporter assays to analyze the Imd, Toll and JAK/STAT pathways, and dsRNA treatments were carried out as described earlier [29,30].

**Drosophila stocks**

The *edin* RNAi line (stock #14289) was obtained from VDRC and the C564-GAL4 flies were a kind gift from from Prof. Bruno Lemaitre (Global Health Institute, EPFL, Switzerland). CG2185 transgenic flies were generated by microinjecting the pUAST-edin construct to the Rel<sup>20</sup> background in the Umea Fly and Worm Transgene Facility. The genotype of the *edin* overexpression fly line is *w*;UAS-*edin*,Rel<sup>20</sup>.

**Lifespan experiments**

*UAS-edin* flies were crossed with C564-GAL4, Actin5C-GAL4/G0 and Dr{\textit{d}}oughtless-GAL4 driver flies. Rel<sup>20</sup> crossed with driver flies were used as a control. The lifespan of the offspring of the crosses was monitored at +25°C. Flies were moved to vials containing 5 ml of fresh fly food twice a week and their survival was monitored. Males and females were kept in separate vials, 10 to 20 flies per vial.

**Infection experiments**

Infections were carried out by pricking one week-old healthy *D. melanogaster* with a thin tungsten needle dipped in a concentrated pellet of either *Escherichia coli*, *Enterobacter cloacae* (strain 12), *Enterococcus faecalis* or *Listeria monocytogenes* (strain 10403S) which were grown overnight on culture plates.

**RNA extraction from flies**

Quadruplicates of five flies (2 females and 3 males) were snap frozen on dry ice 0 h, 1 h, 4 h or 8 h post-infection. Flies were homogenized in TRIzol reagent (Bioline, London, UK) and total RNAs were extracted according to the manufacturer’s instructions.

**Statistical analysis**

Statistical analyses of results were carried out using one-way ANOVA. For survival experiments, Log Rank analysis was carried out and p<0.05 was considered to be significant.

**Flow cytometry**

The amount of cell-associated microbes was analyzed using flow cytometry as described earlier [20].

**Binding assay**

The binding assay for Edin was carried out essentially as described earlier [20] with minor modifications. In brief, S2 cells were seeded onto 24-well plates and transfected with 0.5 μg of pMT-[*edin*-V5 plasmid or an empty pMT-V5 plasmid. CuSO<sub>4</sub> was added 48 h later to a final concentration of 500 μM. Cells were harvested the next day and the supernatant was collected. 1 ml of Schneider medium supplemented with 10% FBS. 50 μl of *E. coli* and *S. aureus* suspension were added to the wells 24 h after CuSO<sub>4</sub> was incubated for 2 h at +25°C. Serial dilutions of the bacterial suspensions were made in sterile water. 20 μl droplets of each dilution were pipetted on LB (*E. coli*) or BHI (*S. aureus*) agar plates, the plates incubated overnight at +37°C and the bacterial colonies counted.

**Functional Characterization of Edin**

*Edin* was induced 48 h later by adding CuSO<sub>4</sub> to a final concentration of 500 μM. *Edin* plasmid or an empty plasmid. Expression of the plasmid was used as a positive control for Gram-positive and Gram-negative strains.

Western blotting

Cells were transfected with 0.5 μg of pMT-[*edin*-V5. Cells were harvested, pelleted and lyzed 24 h after addition of CuSO<sub>4</sub>. 25 μg of cell lysate and supernatant were electrophoresed in NuPAGE 12% Bis-Tris gel (Invitrogen Life Technologies), blotted on a nitrocellulose membrane, and detected by Western blotting using mouse anti-V5 primary Ab (Invitrogen/Life Technologies) and goat anti-mouse Ab HRP conjugates (Molecular Probes) together with ECL Plus Western blotting detection system (GE Healthcare Life Sciences, Uppsala, Sweden).

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Colonies forming unit assay

Edin-V5 expressed in S2 cells: S2 cells in 48-well plates in an antibiotic-free medium were transfected with 0.5 μg of pMT-[*edin*-V5 plasmid or an empty plasmid. Expression of the plasmid was induced 48 h later by adding CuSO<sub>4</sub> to a final concentration of 300 μM. 100 μl of overnight grown bacterial suspension (OD<sub>600nm</sub> = 0.33, ~1*10<sup>6</sup> bacteria/ml) was centrifuged and resuspended in 1 ml of Schneider medium supplemented with 10% FBS. 50 μl of *E. coli* and *S. aureus* suspension were added to the wells 24 h after CuSO<sub>4</sub> and incubated for 2 h at +25°C. Serial dilutions of the bacterial suspensions were made in sterile water. 20 μl droplets of each dilution were pipetted on LB (*E. coli*) or BHI (*S. aureus*) agar plates, the plates incubated overnight at +37°C and the bacterial colonies counted.

Synthetic forms of Edin: An overnight grown bacterial suspension (OD<sub>600nm</sub> = 0.33, ~1*10<sup>6</sup> bacteria/ml) was centrifuged and washed five times with 1 ml of PBS. To detach the bound Edin from the nitrocellulose membrane as described above.
Acknowledgments

We thank other members of our laboratory, Prof. Shoichiro Kurata for the L. monocytogenes strain and VDRC for the edn RNAi line. We are also grateful to Ingrid Dacklin in Umeå Fly and Worm Transgene Facility for performing microinjections to fly embryos. The fly work was done at University of Tampere Drosophila Core Facility funded by Biocenter Finland. All authors approved the final version of the text.

References


Author Contributions

Conceived and designed the experiments: LMV AK JU BW DH SV MR. Performed the experiments: LMV AK MR JU BW DH. Analyzed the data: LMV AK MK JU DH SV MR. Wrote the paper: LMV AK JU BW DH SV MR.